CSF Preparation (15-20ml total)

Step One ➢ Collect initial 1-2 ml CSF for local lab testing.
➢ If bloody, collect CSF until cleared of blood.
➢ If not bloody, transfer into the yellow-capped polypropylene cryovial. Send to local lab for cell count.

Step Two ➢ Place samples upright on wet ice until centrifugation begins.

Step Three ➢ Collect 15-20 ml total (including the 1-2 ml for local lab testing) into two 15 ml polypropylene conical tubes.

Step Four ➢ Within 15 mins of collection, centrifuge samples at 4°C at 2000 x g for 10 minutes.

Step Five ➢ Using a clean transfer pipette, transfer all CSF into a new 50 ml polypropylene conical tube, leaving the pellet in the bottom.
➢ Gently invert the 50 ml conical tube 3-4 times to mix the sample.
➢ Aliquot 1.5 ml CSF into each polypropylene cryovial.
➢ Store CSF aliquots upright at -80°C until shipment.
➢ Spin, aliquot, and freeze aliquots within 2 hours of collection.

Step Six ➢ Label polypropylene cryovials with specimen labels prior to collection.
➢ Pre-chill all polypropylene cryovials on wet ice.
➢ Gravity method for collection is preferred!