A Phase 2 Randomized Double-Blind Placebo-Controlled Trial to Evaluate the Efficacy and Safety of BHV-4157 in Patients with Mild to Moderate Alzheimer’s Disease (T2 PROTECT AD) & The National Centralized Repository for Alzheimer’s Disease and Related Dementias (NCRAD)

Biofluids Collection Training Slides
Training Overview

• GUIDs
• Specimen Collection Schedule
• Kit Request Module
• Specimen Labels
• Handling/Processing Study Specimens
• Sample Shipping
• NCRAD Website
• Questions
Contact Information

• Questions?

Please contact NCRAD Coordinators at:
  • Phone: 317-278-1228 or 1-800-526-2839
  • E-mail: mipetkov@iu.edu or alzstudy@iu.edu
  • Website: www.ncrad.org
Globally Unique Identifier (GUID)

• The GUID is a subject ID that allows researchers to share data specific to a study participant, without exposing personally identifiable information

• A GUID is made up of random alpha-numeric characters and does not include any PHI in the identifier
Globally Unique Identifier (GUID)

1. Create an account: https://bricsguid.nia.nih.gov/portal/jsp/login.jsp
2. Once you have an account, go to the GUID Tool – Create GUID
3. To open the ‘Launch GUID Tool’ you will need to have Java installed on your device
4. When the GUID Tool is open, you will need all of the following information
   • Complete legal given (first)name of participant at birth
   • The participant’s middle name, if applicable
   • Complete legal family (last) name of subject at birth
   • Day of birth
   • Month of birth
   • Year of birth
   • Name of city/municipality in which subject was born
   • Country of birth
## Specimen Collection Schedule

<table>
<thead>
<tr>
<th>Visit</th>
<th>Collection Container</th>
<th>Specimen Type</th>
<th>Container Received</th>
</tr>
</thead>
<tbody>
<tr>
<td>Screening</td>
<td>15 ml Sterile Conical tube(s)</td>
<td>CSF</td>
<td>2.0 ml Cryovials</td>
</tr>
<tr>
<td>Baseline</td>
<td>6 ml Serum (Red-Top) Blood Collection Tube</td>
<td>Serum</td>
<td>2.0 ml Cryovials</td>
</tr>
<tr>
<td></td>
<td>6 ml EDTA (Lavender-Top) Blood Collection Tube</td>
<td>Plasma-PK</td>
<td>2.0 ml Cryovials</td>
</tr>
<tr>
<td></td>
<td>10 ml EDTA (Lavender-Top) Blood Collection Tube</td>
<td>Plasma</td>
<td>2.0 ml Cryovials</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Buffy Coat</td>
<td>2.0 ml Cryovials</td>
</tr>
<tr>
<td>Weeks 4, 8, &amp; 12</td>
<td>6 ml EDTA (Lavender-Top) Blood Collection Tube</td>
<td>Plasma-PK</td>
<td>2.0 ml Cryovials</td>
</tr>
<tr>
<td></td>
<td>6 ml Serum (Red-Top) Blood Collection Tube</td>
<td>Serum</td>
<td>2.0 ml Cryovials</td>
</tr>
<tr>
<td></td>
<td>6 ml EDTA (Lavender-Top) Blood Collection Tube</td>
<td>Plasma-PK</td>
<td>2.0 ml Cryovials</td>
</tr>
<tr>
<td></td>
<td>Sterile Containers</td>
<td>CSF</td>
<td>2.0 ml Cryovials</td>
</tr>
</tbody>
</table>
## Specimen Collection Schedule, cont’d

<table>
<thead>
<tr>
<th>Visit</th>
<th>Collection Container</th>
<th>Specimen Type</th>
<th>Container Received</th>
</tr>
</thead>
<tbody>
<tr>
<td>Week 48</td>
<td>6 ml Serum (Red-Top) Blood Collection Tube</td>
<td>Serum</td>
<td>2.0 ml Cryovials</td>
</tr>
<tr>
<td></td>
<td>6 ml EDTA (Lavender-Top) Blood Collection Tube</td>
<td>Plasma-PK</td>
<td>2.0 ml Cryovials</td>
</tr>
<tr>
<td></td>
<td>10 ml EDTA (Lavender-Top) Blood Collection Tube</td>
<td>Plasma</td>
<td>2.0 ml Cryovials</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Buffy Coat</td>
<td>2.0 ml Cryovials</td>
</tr>
<tr>
<td></td>
<td>15 ml Sterile Conical Tubes</td>
<td>CSF</td>
<td>2.0 ml Cryovials</td>
</tr>
</tbody>
</table>
Kit Request Module

http://kits.iu.edu/t2
Kit Request Module

- An initial stock of kits will be delivered prior to the designated site specific start date.

- Kits and individual supplies available to order:
  - Blood Collection Kits:
    - Baseline, Week 4, Week 8, Week 12, Week 24, Week 48
  - CSF Kit
  - LP 22 Gauge Kit
  - LP 24 Gauge Kit
  - Blood Supplemental Kit
  - CSF Supplemental Kit
  - Frozen Shipping Kit
  - Individual Supplies
Kit Request Module

1. Choose your site from the drop down list.
2. The coordinator name and contact information will populate.
3. Verify that this information is correct.
Kit Request Module: Kit Selection

• Indicate the quantity needed of each kit.

• Once selected, kit components of the chosen kit will appear at the bottom of the screen (Pictured)

• **Note: You can order more than one type of kit in a single kit request**
Kit Request Module: Kit Selection

• For individual supplies, select the ones needed and specify quantities below.
• Click “Submit” to turn in your request.
• The IU staff will notify you that your request has been received and address any issues.
Kit Request Module

• Each site is responsible for ordering kits and maintaining supplies on site for their scheduled participants.

• To order kits, sites will use the Indiana University online kit ordering module: http://www.kits.iu.edu/t2

• Allow around 2 weeks for your order to be processed and delivered.
Specimen Labels
Specimen Labels

• Label type summary:
  • Kit Number Labels
  • Site and ADCS ID Labels
  • Collection and Aliquot Tube Labels
    • Differ by specimen type

• All labels are provided in the kits
Specimen Labels: Kit Number Labels

• Used to track patient samples and provide quality assurance
• Will be placed on:
  • Biological Sample and Shipment Notification Form
  • CSF Sample and Shipment Notification Form
    • CSF samples will have a different kit number than blood samples
  • Outside of cryobox(es) that houses aliquot tubes during storage and shipment
Specimen Labels: Site and ADCS ID Labels

• Subjects will be identified by their site ID and ADCS ID

• Sites will be responsible for handwriting the IDs on the provided labels
  • Fill in labels prior to adhering to tubes
  • Must use fine-point marker
  • Each site will receive a 4-pack of markers in initial kit supply

• Labels will be placed on all collection tubes:
  • Serum Red Top Tube (6 ml)
  • EDTA Lavender Top Tube (6 ml)
  • EDTA Lavender Top Tube (10 ml)
Specimen Labels: Collection & Aliquot Tube Labels

- Specimen Number (Assigned by NCRAD)
- Study Name
- Sample Type (Plasma, Buffy Coat, Serum, Plasma-PK or CSF)
- Kit # (Assigned by NCRAD and unique to the subject and visit)
Specimen Labels: Blood Collection Tubes

- All collection tubes will have two labels:
  - Aliquot and Collection Tube Label
  - Site and ADCS ID Label

![Label 1:](image1)
![Label 2:](image2)

- **Site:** 
- **ADCS ID:**

![Image of collection tubes with labels](image3)

*Place barcode near top*
Specimen Labels: Aliquot Tube Labels (Plasma, Buffy Coat, Serum, Plasma-PK, and CSF)

- Use only the Collection and Aliquot Tube label
- Place barcode near the cap
Specimen Labels: Labeling Biologic Samples

• Label all collection and aliquot tubes **before** cooling, collecting, processing or freezing samples.

• Label only **1** subject’s tubes at a time to avoid mix-ups.

• Wrap the label around the tube **horizontally**. Label position is important for **all** tube types.

• Make sure the label is completely adhered by rolling between your fingers.
Handling/Processing Study Specimens
Site Required Equipment

- **Blood Collection/Safety Equipment:**
  1. Personal Protective Equipment (PPE)
     - Lab Coat, Safety Glasses
  2. Tourniquet
  3. Alcohol Prep Pad
  4. Gauze Pad
  5. Butterfly Needles
  6. Bandage
  7. Sharps Bin and Lid

- **Processing/Storage Equipment:**
  1. Centrifuge capable of $\geq 2000$ rcf with refrigeration to $4^\circ C$
  2. $-80^\circ C$ Freezer
  3. Wet Ice Bucket
# Blood Collection & Processing: Sample Collection Tube

<table>
<thead>
<tr>
<th>Tube Type</th>
<th>Number of Tubes Drawn (per visit)</th>
<th>Tube Image</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serum (Red-Top) Tube (6 ml)</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>EDTA (Lavender-Top) Tube (6 ml)</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>EDTA (Lavender-Top) Tube (10 ml)</td>
<td>1</td>
<td></td>
</tr>
</tbody>
</table>
Blood Collection & Processing: Draw Order

***Important Note***
In order to ensure the highest quality samples are collected, processed, and stored, it is essential to follow the specific collection, processing, and shipment procedures detailed in the following pages. Please read the following instructions first before collecting any specimens. Have all your supplies and equipment out and prepared prior to drawing blood. Please note that the centrifuge may take 30 minutes to cool, so please plan accordingly.

Draw blood in the following order:

1. Plain Red Top Serum Blood Collection Tube (6 ml)
2. EDTA (Lavender-Top) Blood Collection Tube (6 ml) for PK Analysis
3. EDTA (Lavender-Top) Blood Collection Tube (10 ml) for Buffy Coat and Plasma
Blood Collection & Processing: Aliquot Cryovials & Cap Colors

<table>
<thead>
<tr>
<th>Cap Color</th>
<th>Sample Type</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lavender Cap</td>
<td>Plasma</td>
</tr>
<tr>
<td>Red Cap</td>
<td>Serum</td>
</tr>
<tr>
<td>Orange Cap</td>
<td>CSF</td>
</tr>
<tr>
<td>Yellow Cap</td>
<td>CSF to Local Lab</td>
</tr>
<tr>
<td>Blue Cap</td>
<td>Residual</td>
</tr>
<tr>
<td>Clear Cap</td>
<td>Buffy Coat</td>
</tr>
</tbody>
</table>
Aliquot serum to 1.5 ml
Serum Tube - Serum Collection

Up to 2 cryovials possible:
- 2x 1.5 ml in red cap cryovials OR
- 1x 1.5 ml in red cap & 1x <1.5 ml blue cap cryovial
Aliquot plasma-pk to 0.5 ml
EDTA Tube (6 ml) - Plasma PK Collection

- Plasma
- Buffy Coat
- Red Blood Cells

Up to 6 cryovials possible:
6x 0.5 ml Lavender Top OR
5x 0.5 ml Lavender Top and 1x <0.5 ml Blue Top
Plasma and Buffy Coat Preparation (10ml Lavender-Top Tube)

Step One: 
- Store tubes at room temperature.
- Label tubes with preprinted subject labels prior to blood draw.

Step Two: 
- Collect blood in Plasma Tube allowing blood to flow for 10 seconds and ensuring blood flow has stopped.

Step Three: 
- Immediately after blood draw, invert tubes 8-10 times to mix samples.

Step Four: 
- Place thoroughly mixed tube on wet ice until centrifugation begins.

Step Five: 
- Within 30 minutes of blood draw, centrifuge samples at 2000 xg for 10 minutes at 4°C.
- Samples need to be spun, aliquoted, and frozen within 2 hours from time of collection.

Step Six: 
- Label cryovial tubes with preprinted labels.
- Aliquot 1.5 ml into each purple-cap cryovial tube.
- If residual remains, aliquot to a blue-cap cryovial and note the residual on the Biological Sample and Shipment Notification Form.
- Store plasma aliquots at -80°C until shipment.

Step Seven: 
- Label cryovial tube with preprinted label.
- Using a clean transfer pipette, collect theuffy coat (may have residual plasma and some RBCs included).
- Transfer the buffy coat into the cryovial tube.
- Store buffy coat aliquot at -80°C until shipment.

Aliquot plasma to 1.5 ml.
EDTA Tube (10 ml) – Plasma Collection

- Plasma
- BUFFY COAT
- Red Blood Cells

Up to 4 cryovials possible:
3x 1.5 ml in lavender cap cryovials
1x <1.5 ml blue cap cryovial
EDTA Tube (10 ml) – Buffy Coat Collection

- Plasma
- Buffy Coat
- Red Blood Cells

Buffy Coat Aliquot (Please use CLEAR CAP cryovial)
CSF Collection and Processing

***Important Note***

Fasting prior to the lumbar puncture is at the discretion of the site’s PI. However, efforts should be made to keep the conditions under which an LP is performed (fasting, time of day, etc.) consistent for each LP performed on a participant.
Aliquot CSF to 0.5 ml

Step One:
- Label tubes with pre-printed subject labels prior to collection.
- Pre-chill all cryovials on wet ice.

Step Two:
- Collect initial 1-2 ml (if bloody, collect CSF until cleared of blood) into 15 ml conical tube.
- If not bloody, transfer 1-2 ml into the yellow-cap cryovial.
- Send to local lab for testing.

Step Three:
- Collect another 15 ml CSF into a new 15 ml sterile conical tube.

Step Four:
- Place sample upright on wet ice until centrifugation begins.

Step Five:
- Within 15 minutes of collection, centrifuge sample at 4°C for 10 minutes (no brake).

Step Six:
- Using a clean transfer pipette, aliquot 0.5 ml into the orange-cap cryovials, leaving debris at the bottom of the conical tube.
- If a residual aliquot is created, aliquot into blue-cap cryovial. Document specimen number and volume on CSF Sample Notification Form.
- Store CSF aliquots at -80°C until shipment.
Sample Shipping
Frozen Shipping: Guidelines

• Ship Monday-Wednesday Only

• Hold packaged samples in a -80°C freezer until pickup.

• Batch Samples together
  • 5 Cryoboxes
  • Batch shipping should be performed every 3 months or as a full shipment of specimens accumulates, whichever is sooner.
## Sample Shipment Summary

<table>
<thead>
<tr>
<th>Sample Type</th>
<th>Processing/Aliquoting</th>
<th>Tubes to NCRAD</th>
<th>Ship</th>
</tr>
</thead>
<tbody>
<tr>
<td>Whole blood (Red-Top Serum Tube) for Serum Isolation</td>
<td>1.5 ml aliquots per 2.0 ml cryovials</td>
<td>Up to 2</td>
<td>Frozen (Monday-Wednesday)</td>
</tr>
<tr>
<td>Whole blood (Lavender-Top EDTA 6 ml Tube) for Plasma-PK Analysis</td>
<td>0.5 ml aliquots per 2.0 ml cryovials</td>
<td>Up to 6</td>
<td>Frozen (Monday-Wednesday)</td>
</tr>
<tr>
<td>Whole blood (Lavender-Top EDTA tube) for isolation of plasma and buffy coat</td>
<td>Plasma: 1.5 ml plasma aliquots per 2.0 ml cryovials</td>
<td>Up to 4</td>
<td>Frozen (Monday-Wednesday)</td>
</tr>
<tr>
<td></td>
<td>Buffy Coat: 0.75 ml buffy coat aliquot per 2.0 ml cryovial</td>
<td>1</td>
<td>Frozen (Monday-Wednesday)</td>
</tr>
<tr>
<td>CSF</td>
<td>0.5 ml CSF aliquots per 2.0 ml cryovials</td>
<td>Up to 30</td>
<td>Frozen (Monday-Wednesday)</td>
</tr>
</tbody>
</table>
Frozen Shipping: Cryoboxes

Visits: Screening CSF, Baseline Blood

Include only one subject per box.

Labels:
1: SC CSF
2: BL Serum, Plasma, BC, & Plasma-PK

CSF Aliquot tube for local lab (label not provided)
*not shipped to NCRAD
Frozen Shipping: Cryoboxes

Visits: Week 4 Blood, Week 8 Blood, Week 12 Blood

Labels:
1: Week 4 Plasma- PK
2: Week 8 Plasma-PK
3: Week 12 Plasma-PK

Include only one subject per box.
Frozen Shipping: Cryoboxes

Visits: Week 24 CSF & Blood

Include only one subject per box.

Labels:
1: W24 CSF
2: W24 Serum & Plasma-PK

CSF Aliquot tube for local lab (label not provided)
*not shipped to NCRAD
Frozen Shipping: Cryoboxes

Visit: Week 48 CSF & Blood

Include only one subject per box.

Labels:
1: W48 CSF
2: W48 Plasma, BC, Serum, Plasma-PK

CSF Aliquot tube for local lab (label not provided)
*not shipped to NCRAD*
Frozen Shipping: Cryoboxes

CSF, serum, plasma-pk, plasma, and buffy coat aliquots can be combined in one box. Be sure to adhere the necessary Kit Labels on the lid of the cryobox.

Place cryobox in one Biohazard Bag.
Frozen Shipping: Dry Ice Requirements

- Fully cover the cryoboxes with about 2 inches of dry ice in the provided shipper.
- Each Styrofoam shipper can hold up to 5 cryoboxes
- Each Styrofoam shipper must be completely filled and will contain about 45 lbs (20 kg) of dry ice.
Frozen Shipping: Dry Ice Requirements

Class 9 Dry Ice label should not be covered with other stickers and must be completed or the shipping carrier will reject/return your package!

Repository name & address:
NCRAD
IU School of Medicine
351 W. 10th St
TK-342
Indianapolis, IN 46202

Net weight of dry ice in kg

Your name & address

Dry Ice, __kg.

Shipper's name and address

Consignee Name and Address

Note: 2 lbs. = 1kg.
Shipping Frozen Samples

• Schedule FedEx

• Send Biological Sample and Shipment Notification Form to IU ahead of shipment
  • Email: alzstudy@iu.edu or
  • Fax: 317-321-2003
Shipping Regulations and Training

PLEASE NOTE:

• All study personnel responsible for shipping should be certified in biospecimen shipping.

• It is the responsibility of each site to ensure that the appropriate training has been provided and conducted in regards to IATA shipping.

Please see following slides for resources.
Federal Regulations/Training

- Sites are responsible for ensuring proper training is obtained.
- Current federal and international regulations require anyone directly involved with the shipment of potentially infectious materials and other regulated biological materials (including biological specimens and cultures) be properly trained on pertinent shipping requirements.

- **International Air Transport Association (IATA) Training**

<table>
<thead>
<tr>
<th>DGI Training Center</th>
<th>IATA Training Schools</th>
</tr>
</thead>
<tbody>
<tr>
<td>800-338-2291</td>
<td>North America (514)390-6726</td>
</tr>
<tr>
<td>DGltraining.com</td>
<td>Europe, Africa &amp; Middle East (22) 799 2751</td>
</tr>
<tr>
<td>Provides IATA Certified Air Seminars and online courses</td>
<td>Asia, Australia &amp; the Pacific 65 239 7232</td>
</tr>
<tr>
<td></td>
<td><a href="http://www.iata.org">www.iata.org</a></td>
</tr>
<tr>
<td></td>
<td>Training schools located in 30 countries</td>
</tr>
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</table>

<table>
<thead>
<tr>
<th>SaT Pak Inc.</th>
<th>Aiconsult</th>
</tr>
</thead>
<tbody>
<tr>
<td><a href="http://www.saftpak.com">www.saftpak.com</a></td>
<td>Email: <a href="mailto:Airconsult@wanadoo.fr">Airconsult@wanadoo.fr</a></td>
</tr>
<tr>
<td>Provides dangerous goods training via CD or on-site instruction for North America and Europe</td>
<td><a href="http://www.airconsult-bf.com">www.airconsult-bf.com</a></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Bureau of Dangerous Goods LTD., TIANJIN</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Addr.: No.3 Yingshui road, Nankai district, Tianjin China</td>
<td></td>
</tr>
<tr>
<td>Tel: 022-23495890 83326960 83326854 / Fax: 022-83326959</td>
<td></td>
</tr>
<tr>
<td>Email: <a href="mailto:cdmin@bdg-china.com.cn">cdmin@bdg-china.com.cn</a></td>
<td></td>
</tr>
</tbody>
</table>
UN3373 Biological Substance, Category B Training

- Biological Substance, Category B are specimens being transported for “investigational purposes”
- Recommend: investigator sites document training of category B/dangerous goods
- We recommend establishing a record of your staff’s training and date of instruction
- The training records must be made available upon request by the appropriate national authority
  - Additional information from the Department of Transportation (DOT) can be found on their website [http://hazmat.dot.gov](http://hazmat.dot.gov)
Frozen Shipping: FedEx Airbill

• Airbill must be completed or the shipping carrier will reject/return your package!

Sample shipment to NCRAD will be paid for by the T2 grant at UCSD.
Biological Sample and Shipment Notification Forms

• A copy of the sample form *must* be emailed or faxed to NCRAD prior to the date of sample arrival.
• Please include sample forms in all shipments of frozen samples.
• Email: alzstudy@iu.edu
• Fax: 317-321-2003
Biological Sample and Shipment Notification Form

Form: Blood

- Blood Collection for:
  - Serum
  - Plasma-PK
  - Plasma
  - Buffy Coat

- Send by E-mail or fax prior to shipment, and include a copy in each shipment
Biological Sample and Shipment Notification

Form: CSF

Send by E-mail or Fax prior to shipment, and include a copy in each shipment

**CSF Sample and Shipment Notification Form**

*Please email or fax the form on or prior to the date of shipment.*

<table>
<thead>
<tr>
<th><strong>General Information:</strong></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>FedEx tracking #:</td>
<td></td>
</tr>
<tr>
<td>From:</td>
<td>Date:</td>
</tr>
<tr>
<td>Phone:</td>
<td>Email:</td>
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</tbody>
</table>

<table>
<thead>
<tr>
<th><strong>Study:</strong> T2 PROTECT AD</th>
<th><strong>GUID:</strong></th>
<th><strong>Kit #:</strong></th>
</tr>
</thead>
<tbody>
<tr>
<td>Visit: Screening, Week 24, Week 48</td>
<td></td>
<td>KIT BARCODE</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th><strong>Site ID:</strong></th>
<th><strong>ADCS IND #:</strong></th>
<th><strong>Gauge needle used for LP:</strong></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>22G</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th><strong>Sex:</strong> M F</th>
<th><strong>Year of Birth:</strong></th>
<th><strong>CSF Collected?:</strong></th>
<th></th>
</tr>
</thead>
</table>

<table>
<thead>
<tr>
<th><strong>CSF Collection:</strong></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>5. Date of collection: [MMDDYY]</td>
<td>6. Time of collection: [HHMM]</td>
</tr>
<tr>
<td>7. Last time subject ate: [MMDDYY]</td>
<td>8. Last time subject ate: [HHMM]</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th><strong>CSF Processing:</strong></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Time spin started:</td>
<td>[HHMM]</td>
</tr>
<tr>
<td>Duration of centrifugation: Minutes</td>
<td></td>
</tr>
<tr>
<td>Temp of Centrifuge: °C</td>
<td>Rate of centrifugation: kg</td>
</tr>
<tr>
<td>Total amount of CSF collected: mL</td>
<td></td>
</tr>
<tr>
<td>Time aliquoted:</td>
<td>[HHMM]</td>
</tr>
<tr>
<td>Number of 0.5 mL CSF aliquots created (orange cap):</td>
<td>x 0.5 mL</td>
</tr>
<tr>
<td>If applicable, volume of residual CSF aliquot (less than 0.5 mL in blue cap):</td>
<td>mL</td>
</tr>
<tr>
<td>If applicable, specimen number of residual serum aliquot (last four digits):</td>
<td></td>
</tr>
<tr>
<td>Time frozen:</td>
<td>[HHMM]</td>
</tr>
<tr>
<td>Storage temperature in freezer:</td>
<td>°C</td>
</tr>
</tbody>
</table>

| **Notes:** |  |
NCRAD Website

• Helpful Pages:
  • [https://ncrad.org/holiday_closures.html](https://ncrad.org/holiday_closures.html)

<table>
<thead>
<tr>
<th>Date</th>
<th>Holiday</th>
</tr>
</thead>
<tbody>
<tr>
<td>January 1</td>
<td>New Year’s Day</td>
</tr>
<tr>
<td>3rd Monday in January</td>
<td>Martin Luther King Jr Day</td>
</tr>
<tr>
<td>4th Monday in May</td>
<td>Memorial Day</td>
</tr>
<tr>
<td>July 4</td>
<td>Independence Day (observed)</td>
</tr>
<tr>
<td>1st Monday in September</td>
<td>Labor Day</td>
</tr>
<tr>
<td>4th Thursday in November</td>
<td>Thanksgiving</td>
</tr>
<tr>
<td>4th Friday in November</td>
<td>Friday after Thanksgiving</td>
</tr>
<tr>
<td>December 25</td>
<td>Christmas</td>
</tr>
</tbody>
</table>
NCRAD Website: T2 Protect AD Active Study Page

https://ncrad.org/resource_t2protect.html

*Training videos, manual of procedures, and sample forms are available for reference on the T2 Protect AD Active Study Page.
Contact Information

• Questions?
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